

Detect, Identify, and Characterize Cellular versus Non-cellular Aggregates with the Aura CL

Introduction

Cell, protein, and viral aggregates are critical quality attributes for all biological products.¹ Subvisible biotherapeutic product aggregates indicate low product stability and low shelf life. In addition, these attributes are a crucial indicator of potential immunogenicity for a given biological drug. The FDA suggests that “strategies to minimize aggregate formation should be developed as early as feasible in product development.”¹

Quantifying subvisible particle content in cellular therapies is incredibly challenging since cells themselves are subvisible in nature.² The inherent complexity in the manufacture and supply of CAR-T cell therapies, from collection, viral transduction, expansion, and storage, results in greater potential for the formation of large cell and non-cell aggregates affecting clinical safety and efficacy outcomes.³ The complex nature of cell therapy formulations which can include high protein concentrations of human serum albumin for stability purposes, remnant viral vectors from transfection, and cell debris can facilitate the formation of cell aggregates, negatively impacting the therapeutics’ efficacy. Current analytical techniques make the characterization and differentiation of cell aggregates from other subvisible particles virtually impossible. Flow imaging and cytometry techniques struggle to differentiate cell aggregates from other subvisible particles due to the inherent low refractive index contrast of particles in liquid

matrices, influencing sizing accuracy, undercounting and under sizing cell populations in excess of 50%.⁴

The Aura CL™ is the first system specifically designed to detect, count, characterize, and ID cellular aggregates and subvisible particles for product quality measurements in cell therapy, cell line development, and gene therapy applications. The Aura CL is powered by Fluorescent Membrane Microscopy (FMM) to identify fluorescently labeled cells and particles to produce the most accurate and complete characterization of cell and gene therapy products. Aura CL is a fluidics-free platform, preventing clogging issues seen in other methodologies. It enables high-throughput, high refractive index contrast, and 100% measurement efficiency for suspended cells, protein, and viral aggregates found in cell and gene therapies.

This application note introduces the advanced capabilities of the Aura CL to accurately count, characterize, and differentiate immunotherapeutic cells from other aggregates and subvisible particles. Here, we demonstrate the system’s compatibility with well-established cell and protein aggregate staining protocols for particle ID.

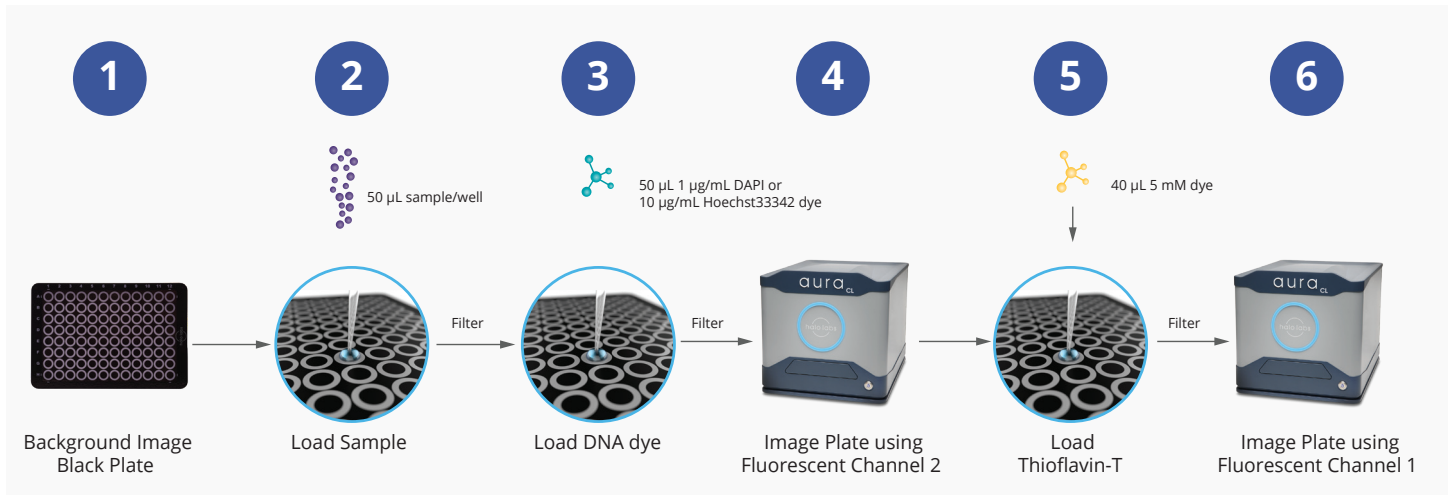


Figure 1: Schematic representation of the DNA and ThT labeling protocols.

Method

Two DNA specific dyes, 4',6-diamidino-2-phenylindole (DAPI) and Hoechst 33342 were specifically chosen and used interchangeably in this application note to distinguish cells from proteins and other particles. Hoechst 33342 is membrane permeable and can stain both live and dead cells while DAPI is cell impermeant at low concentration but can stain live cells at concentrations $>1.0 \mu\text{g/mL}$.

In Figure 1, CAR-T cells were labeled using an “on-membrane” staining protocol. The CAR-T cells were reconstituted at 1×10^5 cells/mL in PBS before 50 µL

was loaded onto a previously background imaged black membrane plate. Samples were vacuumed through and imaged in the Aura CL using brightfield to view all the particles in the sample and side illumination membrane imaging (SIMI) to view non-biologic particles.

50 µL of either 1 µg/mL DAPI or 10 µg/mL Hoechst 33342 was added to the plate and incubated for 10 minutes in the dark to label the cells before the plate was re-imaged using FMM. Nuclear DNA was imaged in the FL2 channel. Samples could also be counterstained on the membrane with Thioflavin T, which specifically labels protein aggregates, and detected using FMM in the FL1 channel.

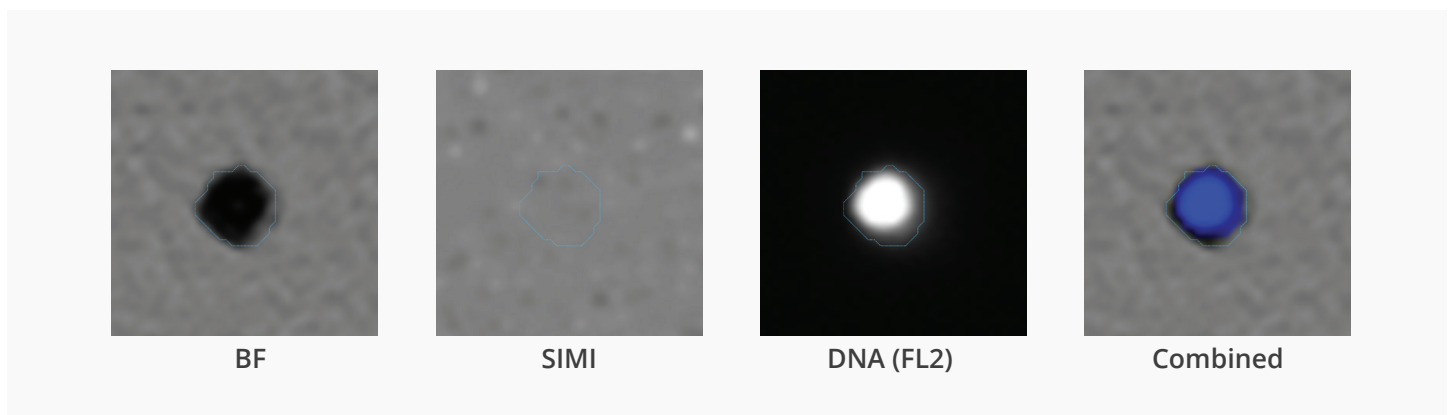


Figure 2: Characteristic morphology of a lymphocyte is observed with a concentric nucleus confirmed through staining with DAPI (FL2 channel, blue).

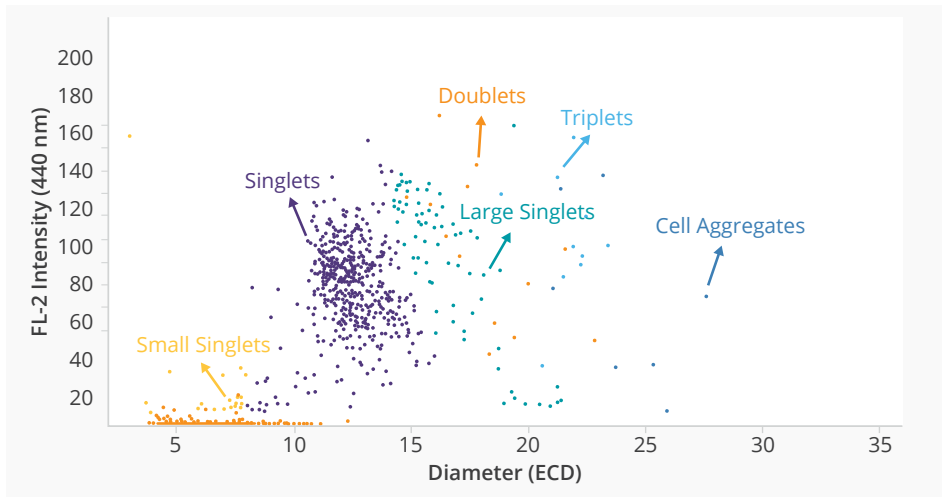


Figure 3: Scatterplot of identified singlets and multimers of CAR-T cells. Particle diameter (X-axis) is plotted against DNA FL2 Intensity (Y-axis). Three types of singlets have been identified, small singlets (yellow), singlets (purple), large singlets (green), as well as doublets (orange), triplets (light blue), and cell aggregates (blue).

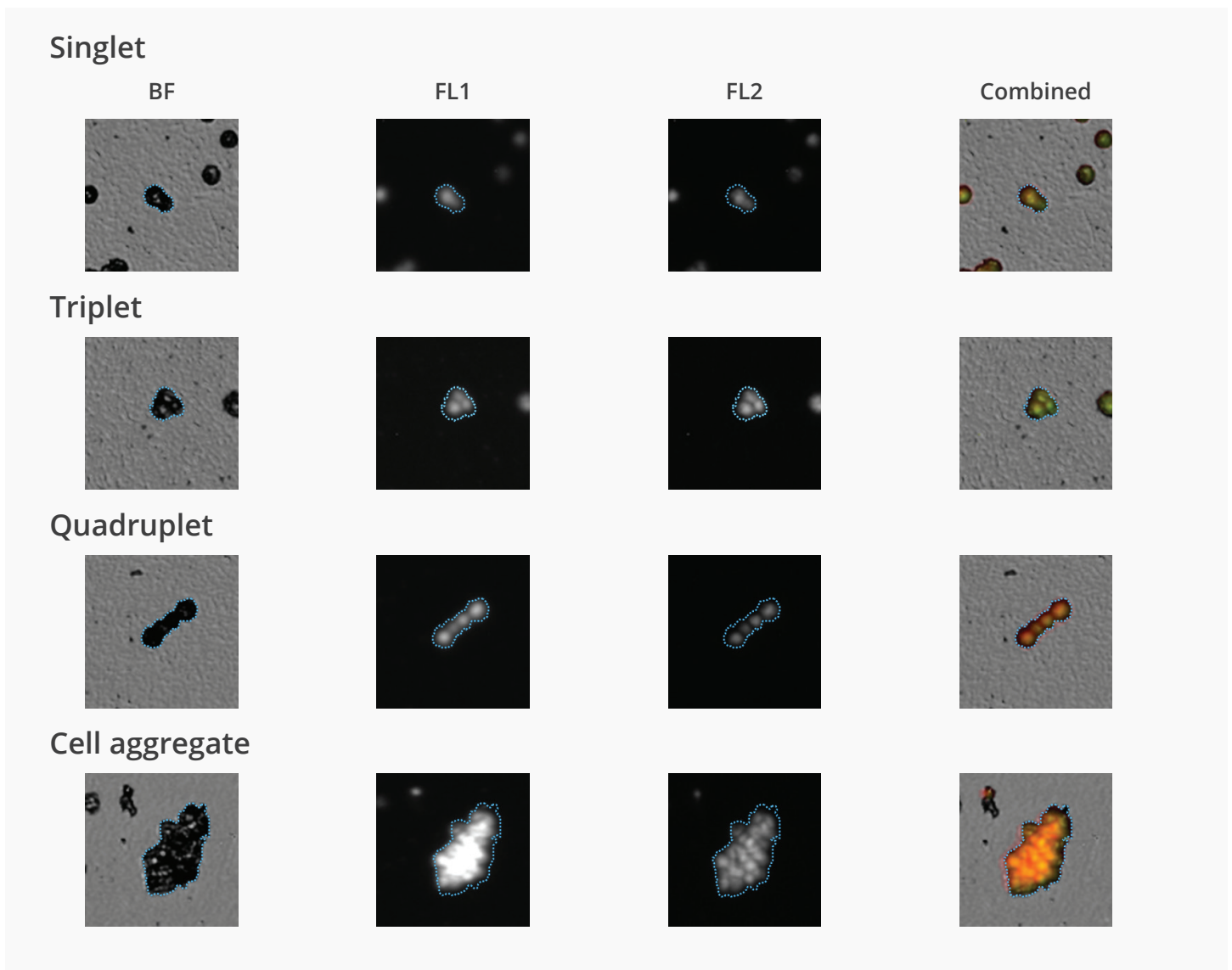


Figure 4: Combined brightfield and fluorescence images from the Aura CL system of cells labeled with a DNA stain (FL2) and ThT for protein (FL1).

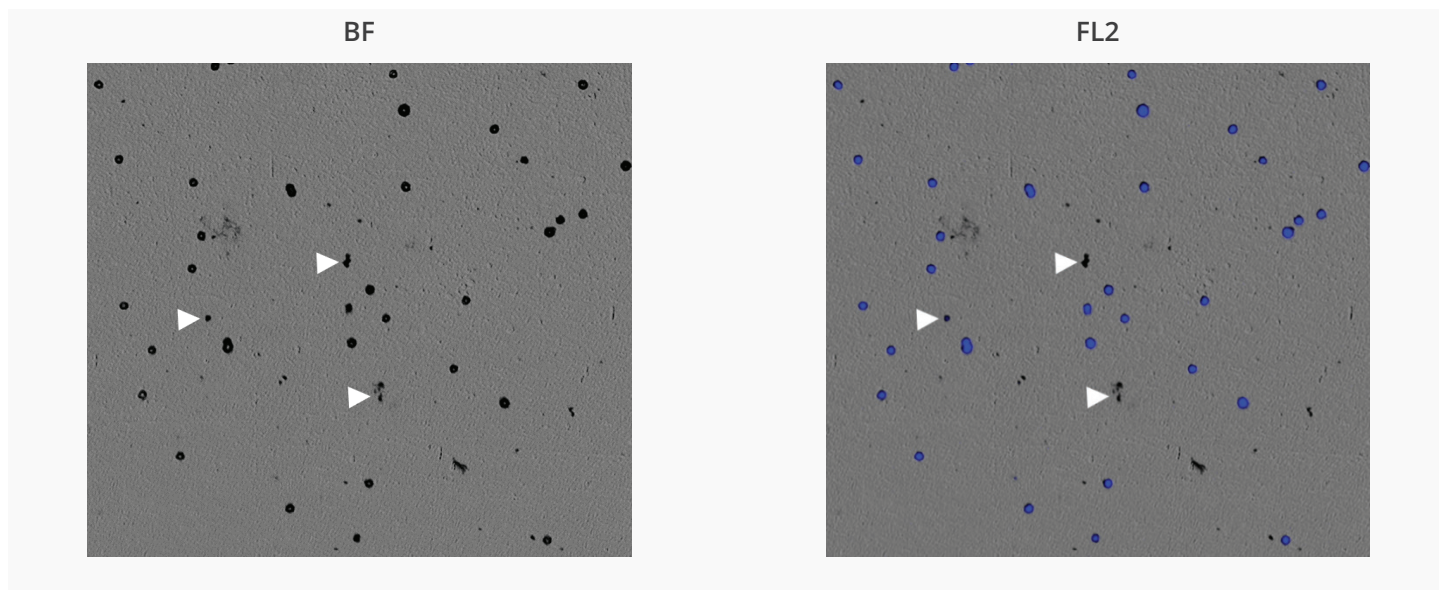


Figure 5: CAR-T cells as imaged by BMI and FMM. Cells can be positively identified using the DNA stain DAPI (blue), highlighting additional non-cellular particles within the sample (arrows).

Results

Singlet and Multimer Analysis

In Figure 2, we show an individual CAR-T cell imaged in brightfield (BMI), SIMI and fluorescence (FL2 channel). With BMI, CAR-T cells demonstrate the distinctive morphology of a lymphocyte, with a concentric condensed nucleus and scanty cytoplasm, with little or no SIMI intensity. In combination with the DNA stain, the CAR-T nucleus is positively identified in the fluorescent channel (FL2).

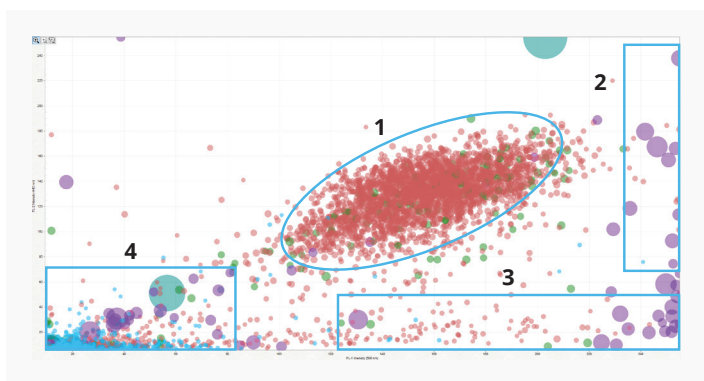


Figure 6: The scatterplot shows individual particles based on their fluorescence, ThT for protein aggregates (FL1 channel, X-axis) and DNA (FL2 channel, Y-axis). Identify cell doublets and triplets (1), cellular aggregates (2), protein aggregates (3), and plastic contaminants (4) in a cell therapeutic sample.

Utilizing the analytical features in Particle Vue 4.0 software, we illustrate how to identify individual cells, doublets, triplets, and larger multimers. In Figure 3, we show a scatterplot of a single well identifying singlets, doublets, triplets, and larger cell aggregates plotted as ECD diameter versus FL2 (DAPI fluorescence) intensity. In this experiment, we expose CAR-T cells to multiple freeze-thaw cycles, leading to an increase in the formation of cell clusters and large cell aggregates. In this data, we observed a striking difference in the uptake of DNA stain in the large cell aggregates, as shown in Figure 4.

Aura CL Specifically Identifies Cells in a Mixed Sample

The Aura CL system images the entire sample and provides information on all particles imaged. In Figure 5, we show a section of a whole well demonstrating the ease by which cells can be distinguished from other subvisible particles (Figure 5, arrows) which are negative for DNA stain.

Different particle populations can be easily viewed using the scatterplot function using Particle Vue software. Cells were labeled with both a DNA nuclei dye and the protein aggregate stain Thioflavin T (ThT) for specific protein identification (Figure 6). Plotting the fluorescent intensity of each particle for each channel easily identifies different particle species. In Figure 6, we observe distinct

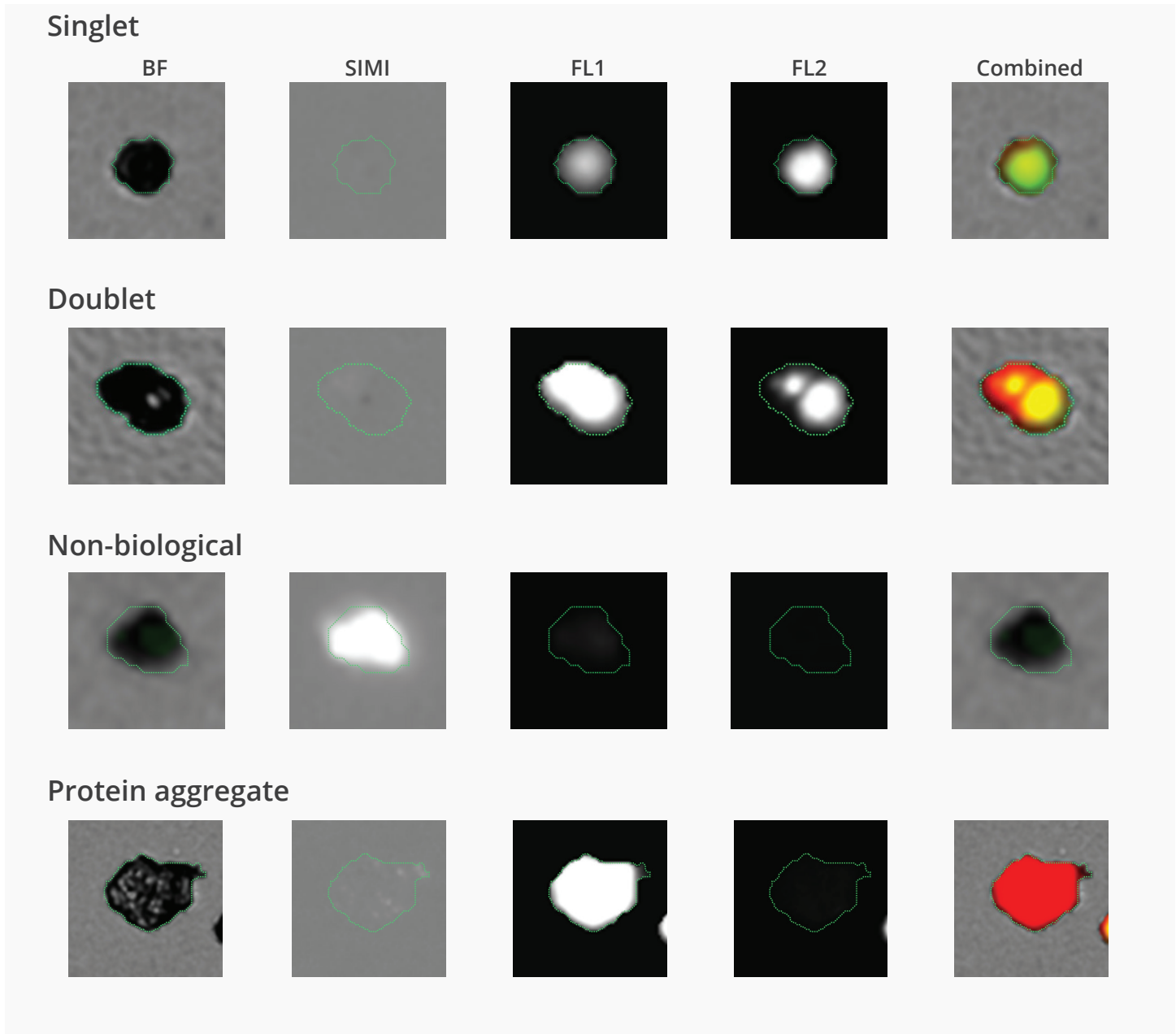



Figure 7: Particles can be identified based upon their fluorescent properties and overall area. Example particles from each quadrant are included here. Singlet and Doublets present dual positive identification for DNA and protein (high FL1 and FL2 intensity), Non-biological aggregate shows particles negative for DNA and protein (Low FL1 and FL2 Intensity), and Protein aggregate shows particles positive for protein only (High FL1 and low FL2 intensity).

populations of cells singlets/doublets/triplets which are positive for both protein (FL1) and DNA (FL2) (circle 1), cell aggregates (square 2) which are positive for both protein (FL1) and DNA (FL2) but are larger in area, protein aggregates positive for protein (FL1) only (box 3), and non-biological particles being negative for both protein and DNA (box 4). The addition of SIMI intensity (side scattering)

can aid in the differentiation of particles, as biological material often resides flat on the membrane and thus exhibit low SIMI. Whereas rigid particles, such as plastic or glass, introduced during the manufacturing process, can exhibit high SIMI intensity profiles (Figure 6 box 4 and Figure 7 Non-biological aggregate), indicating a significant protrusion above the membrane. Conversely, particles

that exhibit low SIMI, and are negative for protein and DNA may be indicative of cell disruption within the therapeutic product (data not presented).

Conclusions

Aura CL is uniquely positioned to fully characterize subvisible particles in cell therapy products and characterize small and large cellular aggregates. Here we showed that using FMM with a combination of DNA and protein aggregate stains allow for the rapid identification of cells, cell aggregates, protein aggregates and non-biological particles in a single, high throughput assay. The assay is also very flexible, enabling analysis of 5 µL of samples to 10 mL of samples, depending on your application and sample availability. The power of FMM is that for first time it is possible to differentiate non-cellular particles in a cell therapy, developing a better understanding of the quality attributes of the product. Compared to standard flow imaging and cytometry techniques, the throughput of FMM is 100x higher, while using best in class particle sizing and counting analysis that has its roots in the well-established membrane microscopy found in USP 788. 

References

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